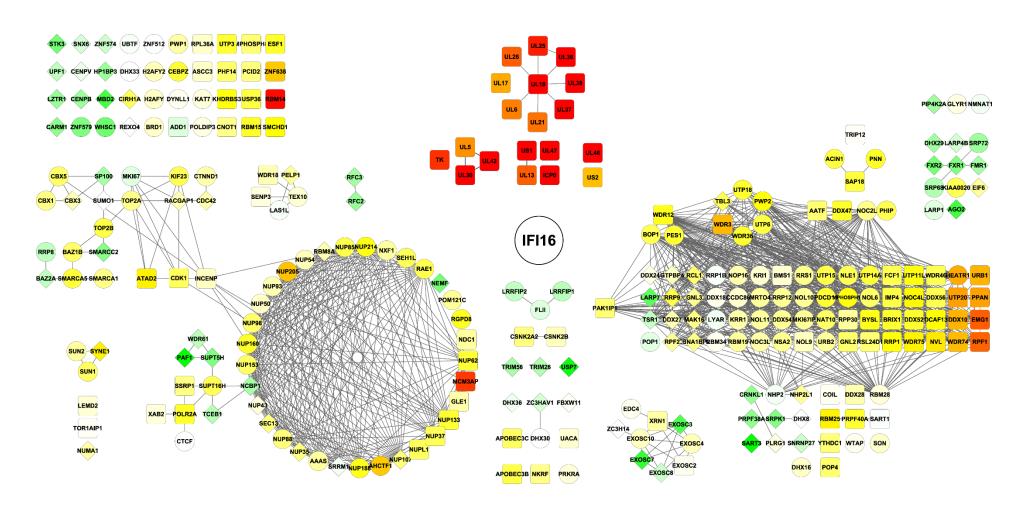
Supplementary Information

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N/A- Provided as separate Excel files

Figure S1.



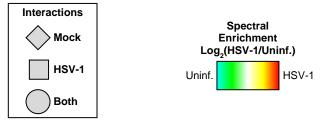
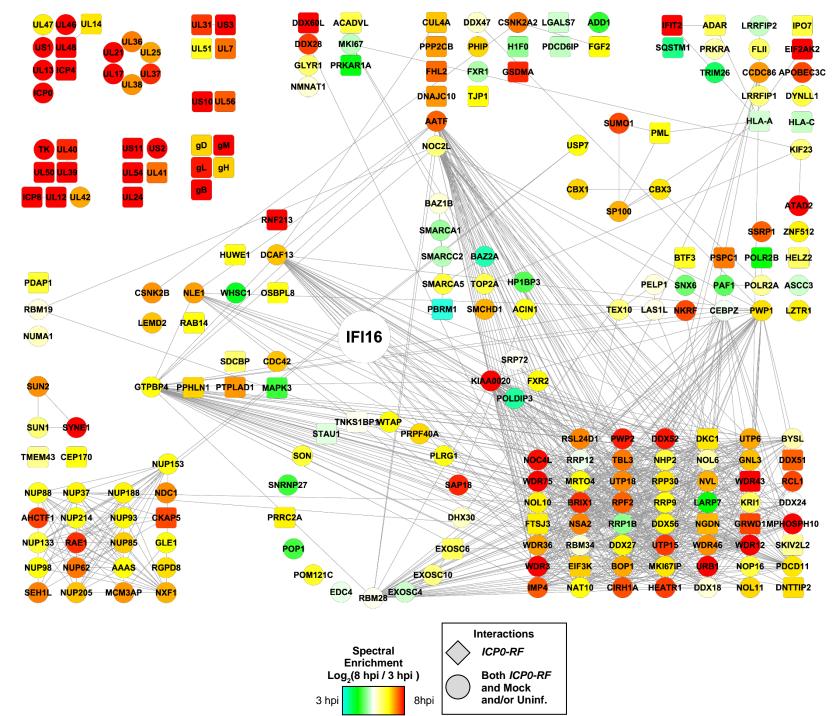
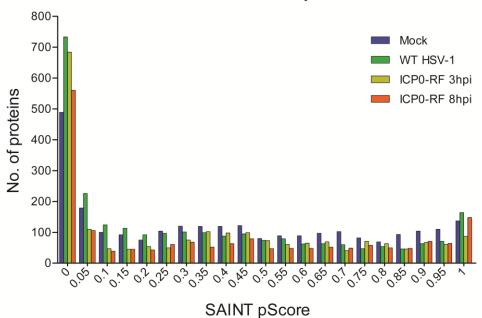


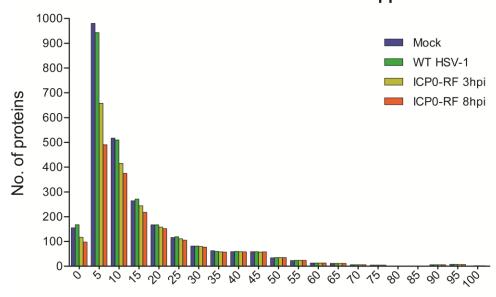
Figure S2.



Distribution of SAINT pScores



Distribution of CRAPome Contaminant Appearance



Frequency of Appearance in CRAPome (%)

Supplementary Figure Legends

Figure S1. Complete interaction network of endogenous IFI16 in uninfected HFF cells and after infection with WT HSV-1. SAINT- and CRAPome-filtered interactions with IFI16 were assembled using STRING and visualized as a network in Cytoscape. Node shape indicates the presence or absence of a protein in other IP conditions. Node colors indicate Log₂-transformed normalized spectral count fold changes between uninfected and WT HSV-1 infected cells.

Figure S2. Complete interaction network of endogenous IFI16 after infection with ICP0-RF at 3 and 8 hpi. SAINT- and CRAPome-filtered interactions with IFI16 were assembled using STRING and visualized as a network in Cytoscape. Node shape indicates the presence or absence of a protein in other IP conditions. Node colors indicate Log₂-transformed normalized spectral count fold changes between 3 and 8 hpi conditions.

Figure S3. Distribution analysis of average SAINT probability scores and CRAPome contaminant frequency for all interactions detected within each IFI16 immunoaffinity purification condition. The distributions of the averages of the top two SAINT scores across three biological replicates for all identified endogenous IFI16 interactions detected in primary fibroblasts for each tested condition are shown (top). Using the 411 total control immunoaffinity isolation experiments submitted to the CRAPome database, percent frequencies of appearance for all identified endogenous IFI16 interactions identified by mass spectrometry within each tested condition were calculated. Their distribution is displayed (bottom).

Supplementary Table Legends

Table S1. IFI16 interactions that passed specificity criteria in uninfected HFFs, or after infection with WT HSV-1. (A) Complete list of IFI16 interactions that passed all the specificity criteria in uninfected HFFs and cells infected with WT HSV-1. (B-C) List of IFI16 interactions that passed SAINT filtering, but not yet the additional filtering based on CRAPome and known cytoskeletal, extracellular, and other abundant sticky proteins; B – uninfected cells; C - cells infected with WT HSV-1. (D) Protein amino acid sequence coverages, illustrated as percentages. (E) Number of unique peptides for all identified proteins.

Table S2. IFI16 interactions that passed specificity criteria in HFFs after infection with *ICP0-RF* at 3 or 8 hpi. (A) Complete list of IFI16 interactions that passed all the specificity criteria in cells infected with ICP0-RF HSV-1. (B-C) List of IFI16 interactions that passed SAINT filtering, but not yet the additional filtering based on CRAPome and known cytoskeletal, extracellular, and other abundant sticky proteins; B - 3 hpi; C - 8 hpi. (D) Protein amino acid sequence coverages, illustrated as percentages. (E) Number of unique peptides for all identified proteins.

Table S3. List of associations excluded as likely non-specific based on CRAPome. Following SAINT filtering, IFI16 interactions were further compared to negative control samples from the web-based CRAPome repository. Proteins with > 20% appearance within the 411 deposited control experiments, as well as common non-specific proteins were filtered out prior to final network analysis.

Table S4. Oligonucleotides for qPCR and shRNA. DNA primer sequences for RT-qPCR of cytokines and oligonucleotide sequences for shRNA constructs.